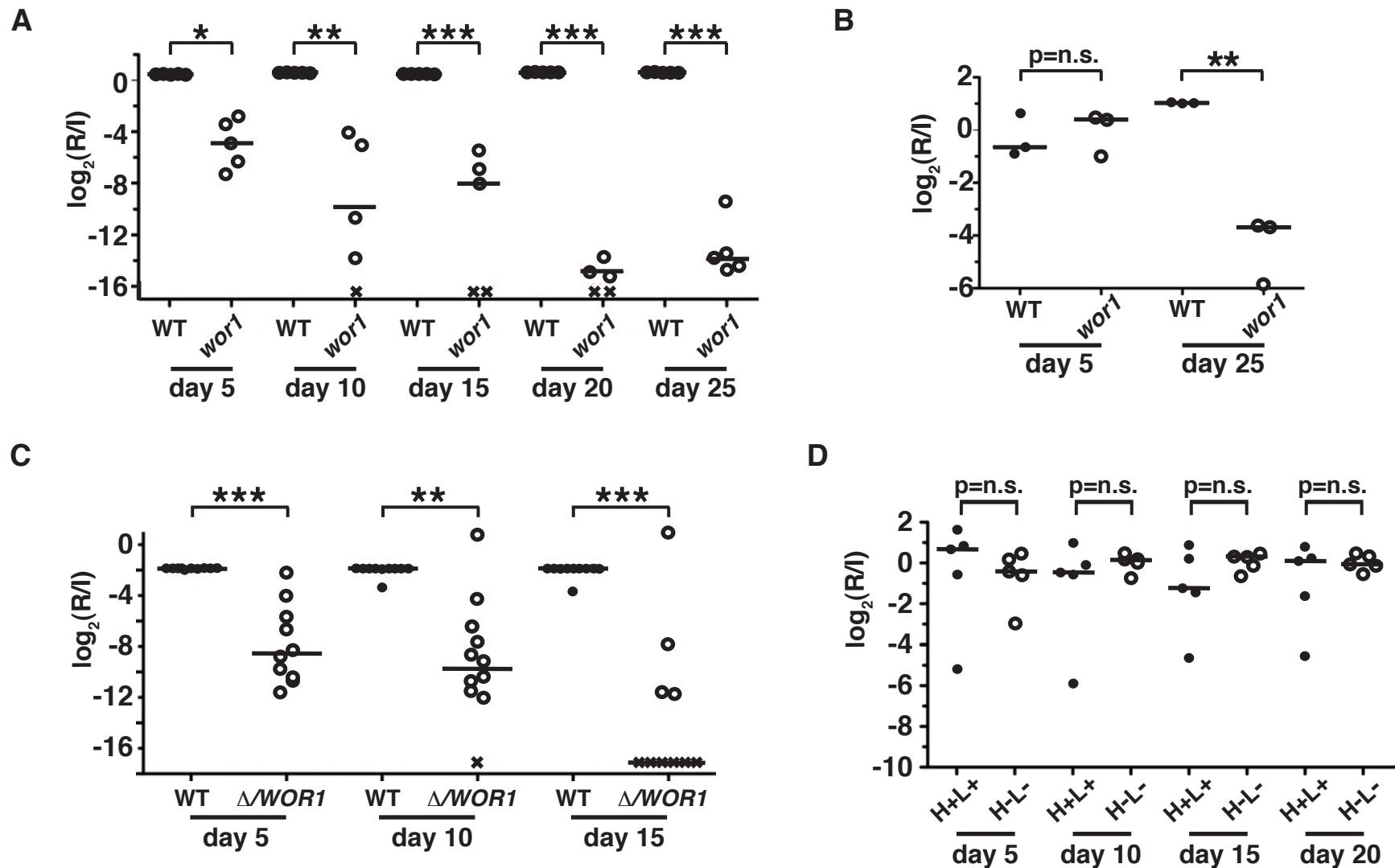


**Passage through the mammalian gut triggers a phenotypic switch that promotes
Candida albicans commensalism**

by Kalyan Pande, Changbin Chen and Suzanne M. Noble

Supplementary Information

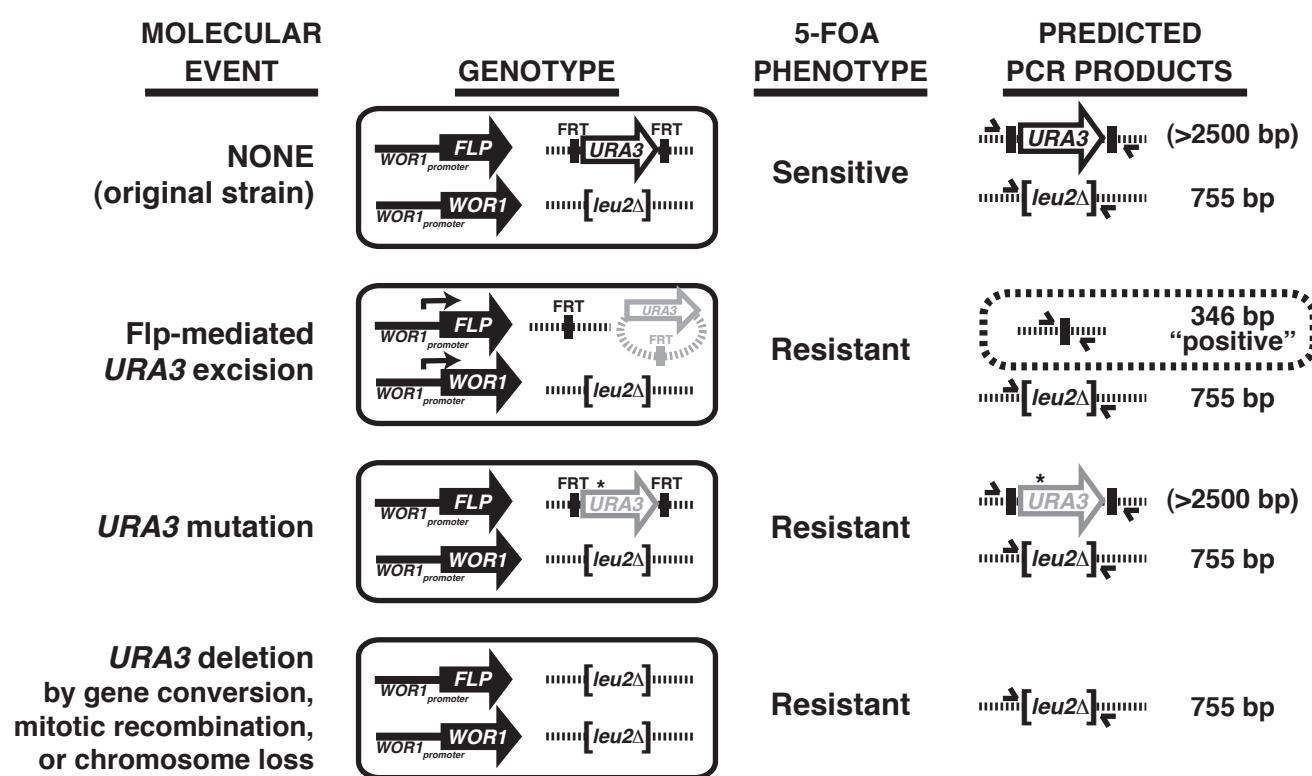
Supplementary Figure 1. *WOR1* is required for wild-type commensal fitness



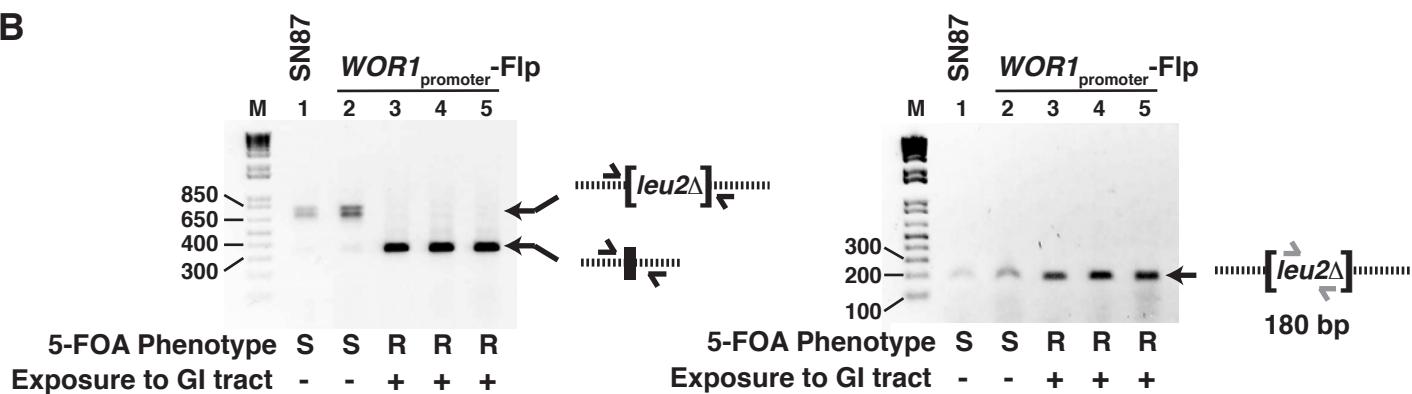
Legend: A) and B) Two independent isolates of *wor1* $\Delta\Delta$ are attenuated in the murine commensal model relative to isogenic wild-type strains. These mutants were generated independently from the *wor1* $\Delta\Delta$ isolate tested in Figure 1C. C) A *wor1* Δ /WOR1 heterozygous deletion mutant is attenuated for commensalism. Note that the haploinsufficiency phenotype can be complemented by increasing WOR1 expression from the remaining allele by means of the TDH3 promoter (*WOR1* OE strain, Figure 3A). D) A His-Leu- double auxotroph exhibits wild-type commensalism. Strain designations are: (A) *wor1* $\Delta\Delta$ (SN1064), WT (SN425); B) *wor1* $\Delta\Delta$ (RZY244), WT (QMY23); C) *wor1* Δ /WOR1 (SN999), WT (SN235); D) His-Leu- auxotroph (SN87), prototroph (SN425). Significance was determined using the t-test: A) *p<0.005, **p<0.001, ***p<0.0001; B) n.s. not significant, **p<0.002; C) **p=0.0002, ***p<0.0001; D) n.s. not significant.

Supplementary Figure 2. Estimation of *WOR1* gene expression in *MTLa/α* cells

A



B



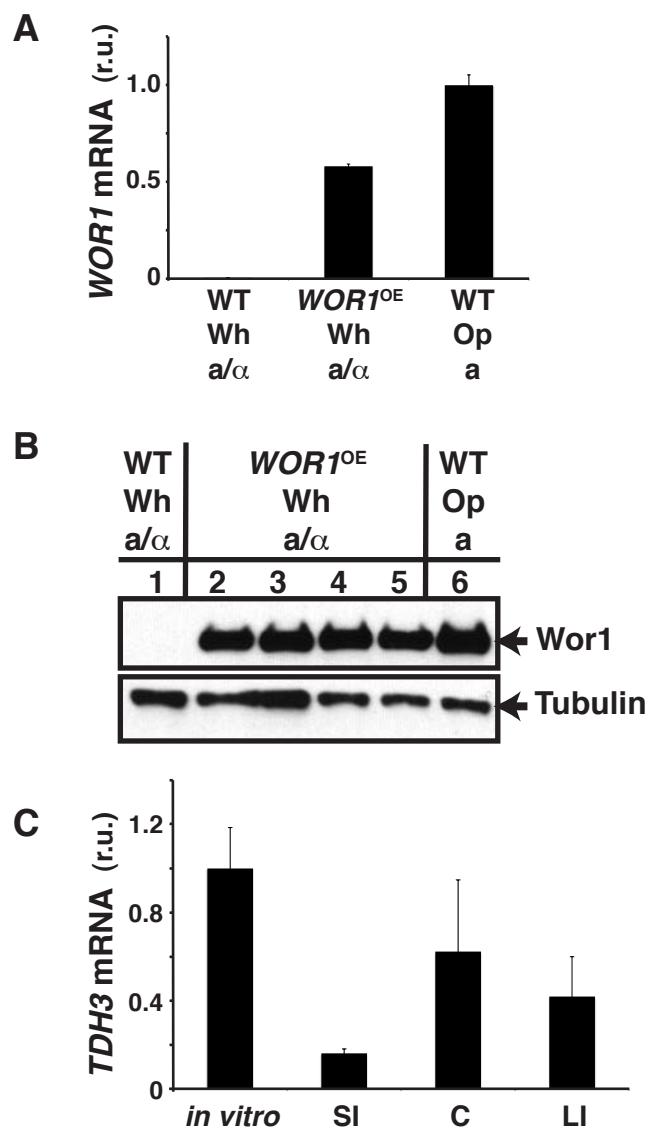
C

	# Cells	# 5-FOA-resistant	# 5-FOA-resistant, PCR-positive	<i>WOR1</i> expression frequency	Standard deviation
<i>in vivo</i>	1.60 x 10e6	3.17 x 10e4	3.14 x 10e4	1.94%	0.55%
<i>in vitro</i>	1.79 x 10e6	47	4	0.0002%	0.0001%

Supplementary Figure 2 Legend: Estimation of *WOR1* gene expression in *MTLa/α* cells.

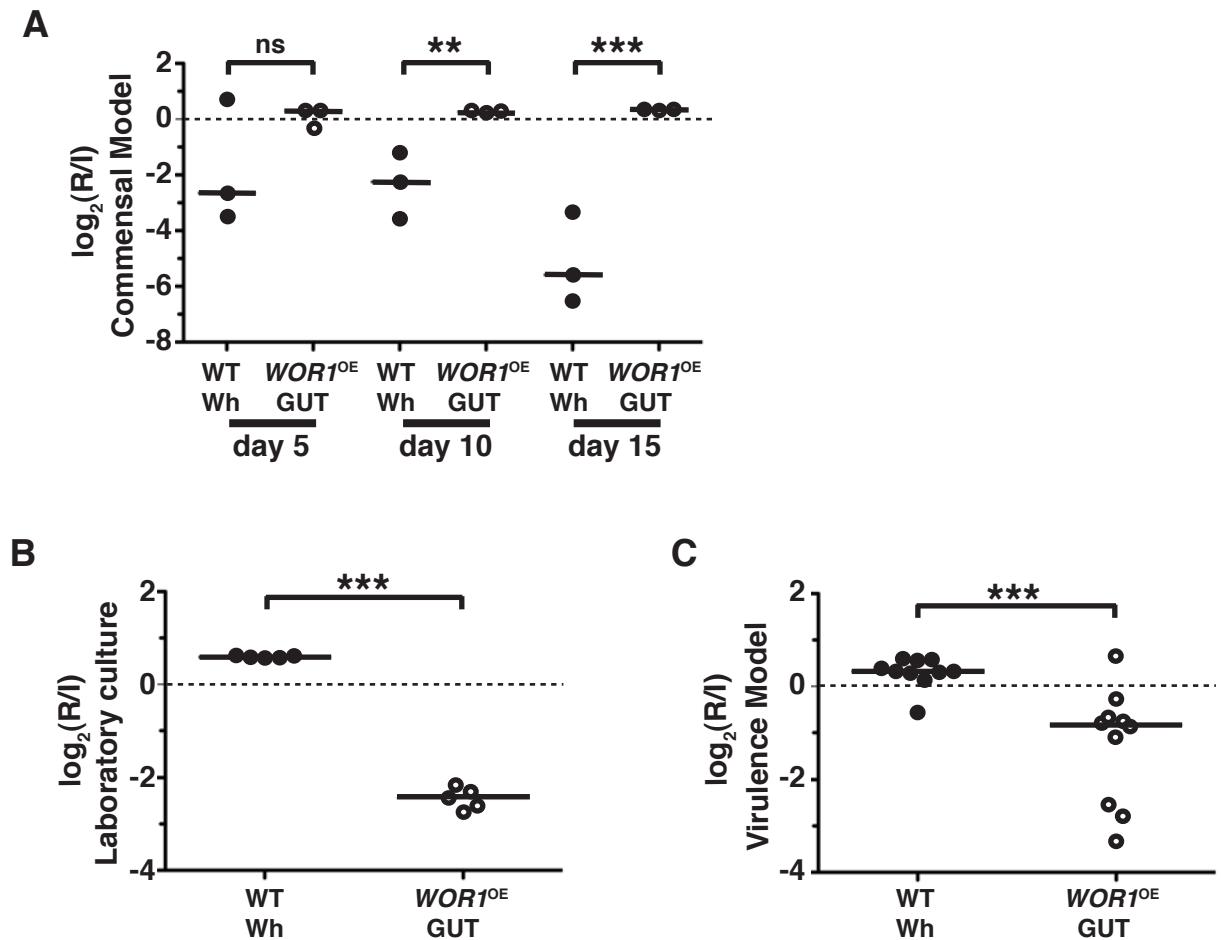
A) Strategy for capturing *WOR1* expression events. The *WOR1*^{promoter}-*FLP* strain (SN1020) has the following genotype: *wor1Δ::WOR1*^{promoter}-*FLP/WOR1*, *leu2Δ::FRT-URA3-FRT/leu2Δ*. Activation of the *WOR1* promoter in this strain results in transcription of *FLP*, followed by Flp-mediated excision of *URA3*, and acquisition of resistance to 5-FOA. Note that 5-FOA-resistance may alternatively arise from mutation of the *URA3* gene or loss of *URA3* through mitotic recombination, gene conversion (via homologous sequences on the other copy of Chromosome 7), or chromosome loss. Colony PCR is performed to screen for products of Flp-mediated recombination, which generate a unique 346 bp product (“positive” test). B) Representative PCR results. On the left, PCR for Flp-mediated events is shown for SN87 (lane 1), the reference strain used to construct *WOR1*^{promoter}-*FLP*; the *WOR1*^{promoter}-*FLP* starting strain (lane 2); and three 5-FOA-resistant isolates of *WOR1*^{promoter}-*FLP* (lanes 3-5) that were recovered from animals. Note that the trans-FRT reaction in Flp-recombinant strains outcompetes the trans- *leu2Δ* reaction, such that only the 346 bp product is apparent in lanes 3-5. On the right, PCR using primers internal to the *leu2Δ* locus confirms that all five strains retain this locus. C) Frequency of Flp-mediated *URA3* excision *in vivo* and *in vitro*. The *WOR1*^{promoter}-*FLP* strain was propagated for three days in the commensal model (n=6 mice) or for 8 generations *in vitro* at 37°C (n=4 cultures). The frequency of cells expressing *WOR1* propagated under a given condition was estimated from the ratio of (5-FOA^R, PCR-positive colonies) / (total number of colonies). Diagnostic PCR was performed on 288 of the 31,700 5-FOA-resistant isolates recovered from mice and on all 47 5-FOA-resistant isolates obtained *in vitro*. 277/278 *in vivo* strains and 4/47 *in vitro* strains tested positive for the 346 bp marker of Flp-mediated recombination. Average values with standard deviations were plotted.

Supplementary Figure 3. Characterization of the *WOR1^{OE}* strain



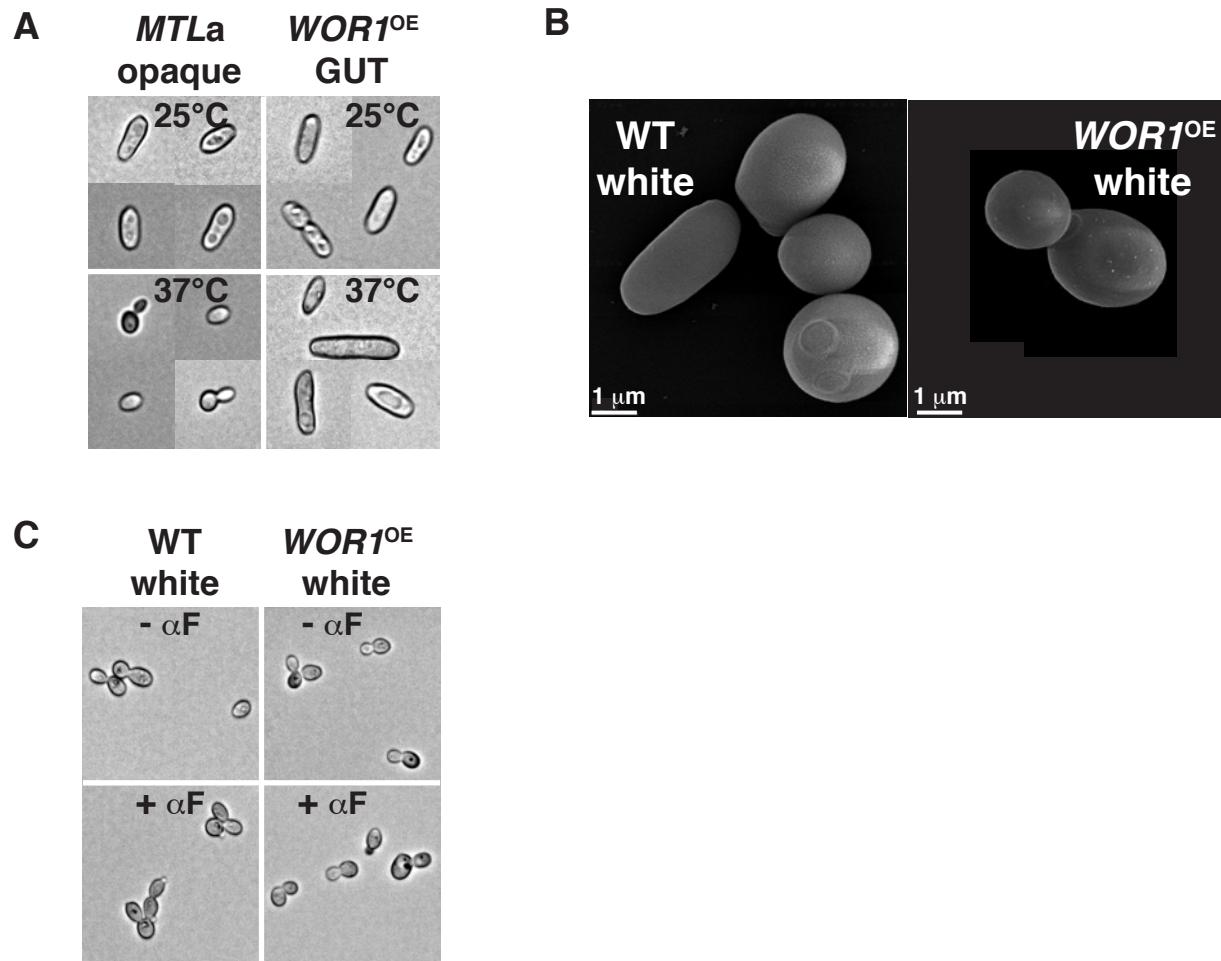
Legend: A) *WOR1* mRNA is highly expressed in the *WOR1^{OE}* (*TDH3_{promoter}-WOR1*) strain. RT-qPCR was performed on three biological replicates each of wild-type white (*MTLa/a*, SN425), *WOR1^{OE}* white (*MTLa/a*, SN928), and opaque (*MTLa*, SN967) strains. Median values are shown, along with standard deviations. Relative to its expression in WT cells, *WOR1* is induced 102-fold in the *WOR1^{OE}* strain and 175-fold in opaque cells. B) Wor1 protein is highly expressed in the *WOR1*-overexpression strain. Immunoblot of Wor1 protein prepared from wild-type white (*MTLa/a*, SN425), *WOR1^{OE}* white (*MTLa/a*, SN828 and three additional isolates), and opaque (*MTLa*, SN967) strains. C) In the mammalian GI tract, the *TDH3* promoter is active but not induced relative to *in vitro*. RT-qPCR of mRNA from wild-type white cells (*MTLa/a*, SN425) after 8 doublings in liquid YEPD medium at 37°C (*in vitro*) or directly recovered from small intestines (SI), ceca, or large intestines (LI) after 10 days in the murine commensal model. Medians with standard deviations are shown for 5 (laboratory culture) or 3 (commensalism model) biological replicates.

Supplementary Figure 4. *MTLa/α* GUT cells display an early fitness advantage in the mammalian GI tract, but are less fit in laboratory culture and in blood-stream infection



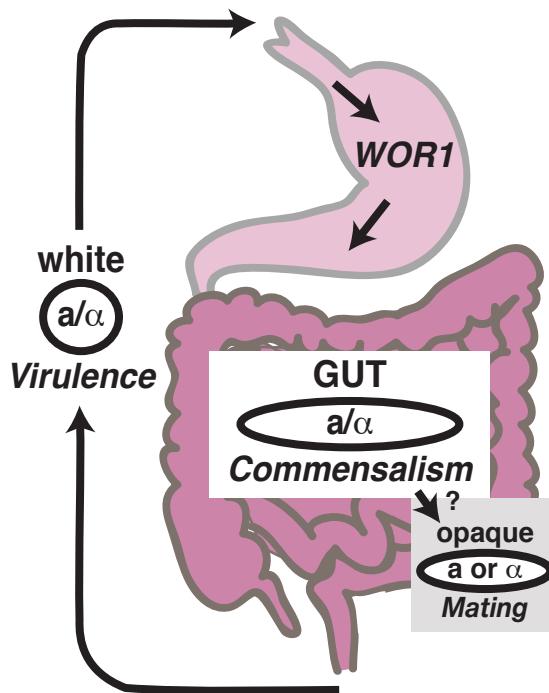
Legend: A) Commensal competition experiment between WT *MTLa/α* white cells (SN425) and *WOR1^{OE}* *MTLa/α* GUT cells (SN1045). Note that, in contrast to the case with white phase *WOR1^{OE}* (Figure 2A), GUT phase cells are highly competitive from the beginning of the time course. ns nonsignificant, ** p<0.02, *** p<0.005. B) *in vitro* competition between the same two strains. Strains were co-cultured for 8 generations in liquid YEPD medium at 37°C. Relative abundances of strains at the beginning (I) and end (R) of the time course were determined by qPCR. *** p<0.0001. C) Competition between the same two strains in a murine model of disseminated disease. Strains were co-infected into BALB/c mice via lateral tail vein, *C. albicans* was recovered from kidneys of moribund animals after a median of 5 days post infection (range 4 to 6 days), and relative abundances in the inoculum (I) and after recovery from kidneys (R) was determined by qPCR.

Supplementary Figure 5. Additional phenotypes of GUT and white cells



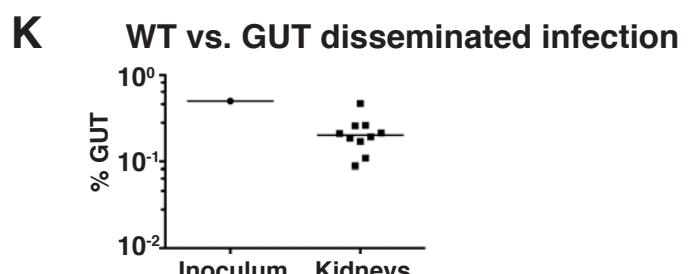
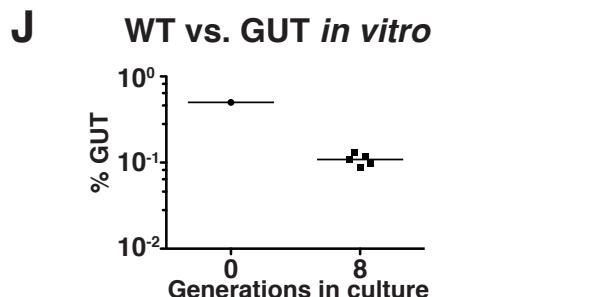
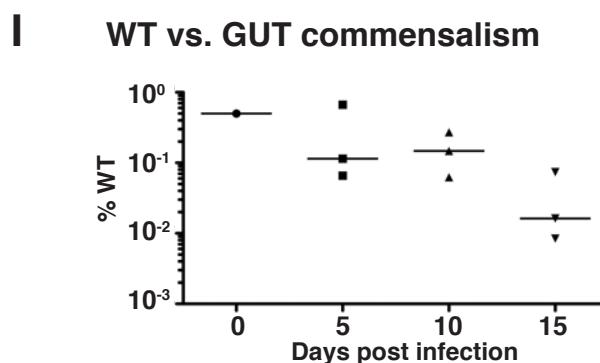
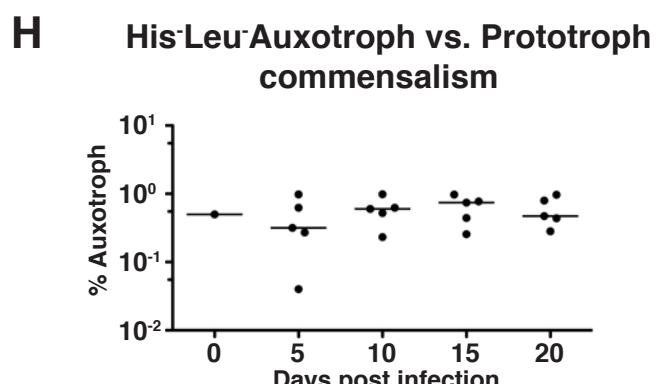
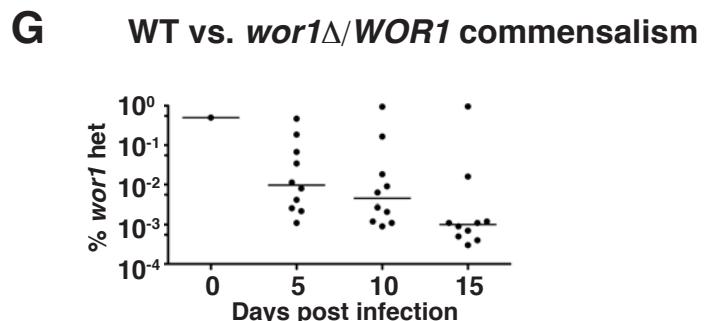
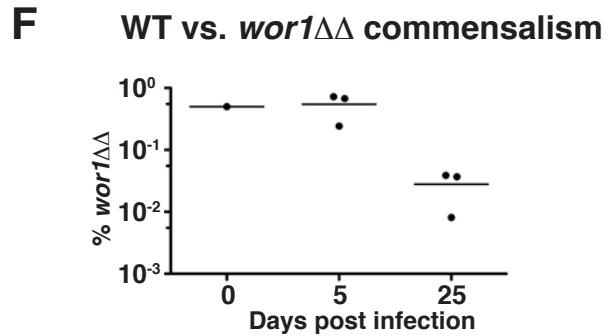
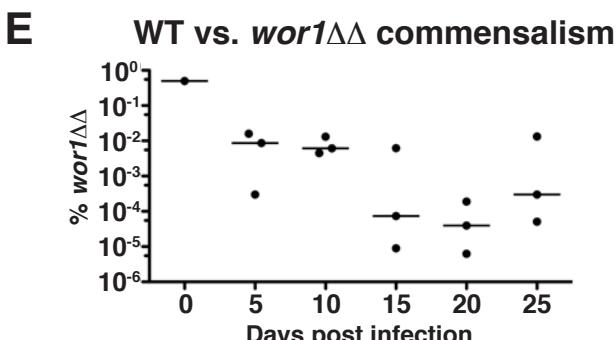
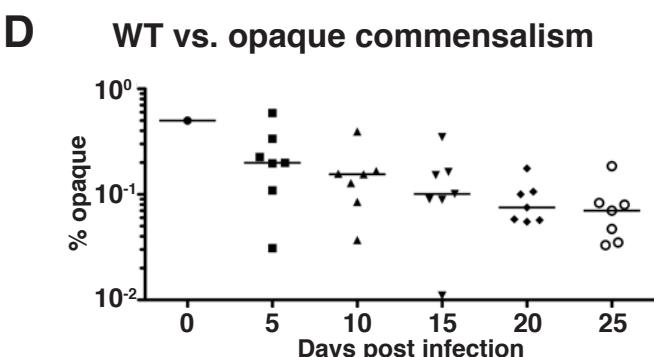
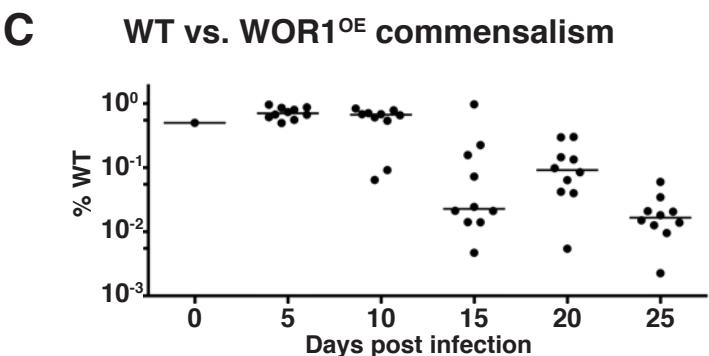
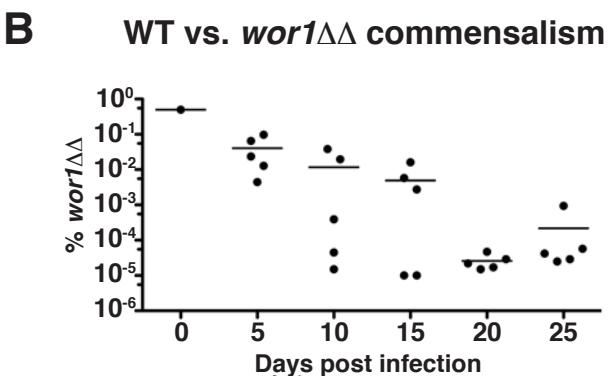
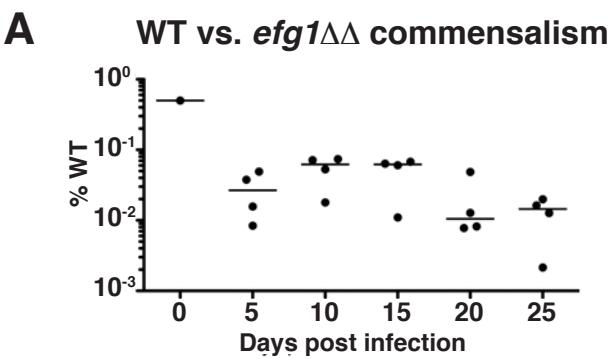
Legend: A) The GUT phenotype is stable at 37°C. After 3 days of incubation on YEPD medium at room temperature (~25°C) or 37°C, *MTLa*/α GUT (SN1045) and *MTLa* opaque (SN967) cells were visualized by light microscopy. B) SEM images of white phase WT (SN425) and *WOR1^{OE}* (SN928) cells, which lack pimple structures. C) White phase *MTLa* (SN966) and *WOR1^{OE}* (SN1044) strains fail to produce germ tubes in response to mating pheromone.

Supplementary Figure 6. Model of phenotypic switching in the host



Legend: According to this model, *MTL* a/α white cells exposed to the mammalian GI tract encounter signals triggering *WOR1* expression and a shift to the GUT state, which is specialized for commensalism. GUT cells exiting this space rapidly convert back to the white state, which is virulent. Additional signals may alternatively trigger GUT cells to undergo loss of one allele of *MTL* (e.g. through loss of Chromosome 5) and conversion to the sexually-competent opaque state.

Supplementary Figure 7



Supplementary Figure 7 Legend: Competition experiments in “% abundance of the less fit strain” format. A) WT (SN250) is outcompeted by *efg1ΔΔ* (SN1011) in the murine commensal model. Plot is derived from the same data as depicted in **Figure 1A**. B) *wor1ΔΔ* (SN881) is outcompeted by WT (SN250) in the murine commensal model. Plot is derived from the same data as depicted in **Figure 1C**. C) WT (SN425) is outcompeted by *WOR1OE* (SN928) in the murine commensal model. Plot is derived from the same data as depicted in **Figure 2A**. D) Opaque cells (SN967) are outcompeted by WT white cells (SN425) in the murine commensal model. Plot is derived from the same data as depicted in **Figure 3C**. E) *wor1ΔΔ* (SN1064) is outcompeted by WT (SN425) in the murine commensal model. Plot is derived from the same data as depicted in **Supplementary Figure 1A**. F) *wor1ΔΔ* (RZY244) is outcompeted by WT (QMY23) in the murine commensal model. Plot is derived from the same data as depicted in **Supplementary Figure 1B**. G) *wor1Δ/WOR1* heterozygous knockout (SN999) is outcompeted by WT (SN235) in the murine commensal model. Plot is derived from the same data as depicted in **Supplementary Figure 1C**. H) A His-Leu- double auxotroph (SN87) is as fit as a prototroph (SN425) in the murine commensal model. Plot is derived from the same data as depicted in **Supplementary Figure 1D**. I) White phase WT (SN425) is outcompeted by GUT cells (SN1045) in the murine commensal model. Plot is derived from the same data as depicted in **Supplementary Figure 4A**. J) GUT cells (SN1045) are outcompeted by white phase WT (SN425) in liquid culture medium. Plot is derived from the same data as depicted in **Supplementary Figure 4B**. K) GUT cells (SN1045) are outcompeted by white phase WT (SN425) in a murine model of disseminated candidiasis. Plot is derived from the same data as depicted in **Supplementary Figure 4C**.

Supplementary Table 1. *MTLa* and *MTLα* Genotypes of White and GUT Cells After Recovery from the Murine Commensal Model

Experiment	Phenotype	<i>MTLa</i> present	<i>MTLα</i> present	Colonies analyzed	% <i>MTLa/α</i>
1	white	44	44	44	100
1	GUT	32	40	40	80
2	white	53	52	53	98
2	GUT	7	8	8	88
3	white	24	24	24	100
3	GUT	24	24	24	100
4	white	24	24	24	100
4	GUT	24	24	24	100
Total	white	145	144	145	99
Total	GUT	87	96	96	91

Supplementary Table 2. Mating Efficiencies of Various Strains

Cross	Strain and Phase		Mating Efficiency (His ⁺ Arg ⁺ /His ⁺ Arg ⁻)		
	Strain 1 (His ⁺ Arg ⁻)	Strain 2 (His ⁻ Arg ⁺)	3 days	5 days	13 Days
1	<i>MTLα</i> opaque	<i>MTLa</i> opaque	3.3 x 10-1	8.0 x 10-1	1.5 x 10-1
2	<i>MTLα/α</i> white	<i>MTLa</i> opaque	<6 x 10-7	<2 x 10-7	<1 x 10-7
3	<i>WOR1^{OE}</i> <i>MTLa/α</i> white	<i>MTLa</i> opaque	4.6 x 10-6	4.2 x 10-6	5.7 x 10-6
4	<i>WOR1^{OE}</i> <i>MTLα/α</i> GUT	<i>MTLa</i> opaque	<2 x 10-7	<3 x 10-7	<2 x 10-7

Supplementary Table 4. Regulated Genes Sets in GUT and opaque cells

Gene Name Standard Name	Description of Gene Product	Cellular Process	Reported Specificity (Wh vs. Op)			RNA Expression Level					Relative Expression		
			Lan et al.	Tsong et al.	Tuch et al.	WT Wh	WOR1 Wh	WOR1 GUT	MTLa Wh	MTLa Op	GUT/ Wh	Op/ Wh	GUT/ Op
UPREGULATED IN GUT AND OPAQUE (VS. WH CONTROLS)													
PXP2 <i>orf19.1655</i>	Putative acyl-CoA oxidase	Lipid Utilization	Op	Op	Op	1.7	7.6	30.0	1.0	47.7	17.8*	47.7*	0.6
ANT1 <i>orf19.6254</i>	Putative adenine nucleotide transmembrane transporter	Lipid Utilization	Op	Op	Op	1.1	2.7	6.8	1.0	5.7	6.4*	5.7*	1.2
FOX2 <i>orf19.1288</i>	3-hydroxyacyl-CoA epimerase	Lipid Utilization	Op			1.2	3.3	5.0	1.0	6.1	4.1*	6.1*	0.8
POX1 <i>orf19.5723</i>	Putative acyl-CoA oxidase	Lipid Utilization	Op		Op	1.1	2.2	4.1	1.0	5.0	3.9*	5.0*	0.8
ECI1 <i>orf19.6445</i>	Ortholog of <i>S. cerevisiae</i> Eci1p, which is involved in fatty acid oxidation	Lipid Utilization			Op	1.1	2.2	3.1	1.0	5.2	2.9*	5.2*	0.6*
POX1-3 <i>orf19.1652</i>	Putative acyl-CoA oxidase	Lipid Utilization				1.0	1.8	3.9	1.0	3.1	3.8*	3.1*	1.3
CAT2 <i>orf19.4591</i>	Major carnitine acetyl transferase	Lipid Utilization		Op		1.1	1.8	2.5	1.0	2.6	2.2*	2.6*	0.9
POT1 <i>orf19.7520</i>	Putative peroxisomal 3-oxoacyl CoA thiolase	Lipid Utilization				1.0	1.9	2.8	1.0	2.7	2.7*	2.7*	1.1
SPS20 <i>orf19.4157</i>	Peroxisomal 2,4-dienoyl-CoA reductase	Lipid Utilization	Op	Op	Op	1.0	1.7	3.0	1.0	5.2	2.9*	5.2*	0.6
NAG1 <i>orf19.2156</i>	Glucosamine-6-phosphate deaminase	NAG Utilization	Op		Op	1.0	4.2	17.7	1.0	10.6	17.4*	10.6*	1.7*
DAC1 <i>orf19.2157</i>	N-acetylglucosamine-6-phosphate (GlcNAcP) deacetylase	NAG Utilization		Op	Op	1.2	2.5	10.0	1.0	5.5	8.3*	5.5*	1.8*
HXK1 <i>orf19.2154</i>	N-acetylglucosamine (GlcNAc) kinase	NAG Utilization			Op	1.0	1.5	2.1	1.0	2.2	2.1*	2.2*	0.9

UPREGULATED IN OPAQUE ONLY (VS. WH CONTROLS)													
SAP2 <i>orf19.3708</i>	Secreted aspartyl proteinase	Protein Utilization		Op	Wh	1.2	1.2	1.3	1.0	4.4	1.1	4.4*	0.3*
SAP3 <i>orf19.6001</i>	Secreted aspartyl proteinase	Protein Utilization	Op	Op		1.0	1.2	1.1	1.0	2.5	1.1	2.4*	0.5*
SAP4 <i>orf19.5716</i>	Secreted aspartyl proteinase	Protein Utilization		Op		1.3	1.7	1.7	1.0	11.9	1.3	11.9*	0.1*
SAP6 <i>orf19.5542</i>	Secreted aspartyl proteinase	Protein Utilization			Wh	1.5	1.7	1.6	1.0	12.0	1.1	12.0*	0.1*
SAP7 <i>orf19.756</i>	Secreted aspartyl proteinase	Protein Utilization				1.9	4.5	1.0	1.9	5.5	0.5	2.9*	0.2*
SAP99 <i>orf19.853</i>	Putative secreted aspartyl protease	Protein Utilization	Op		Op	1.1	1.3	1.4	1.0	7.3	1.3	7.3*	0.2*
DOWNREGULATED IN GUT AND OPAQUE (VS. WH CONTROLS)													
ALS2 <i>orf19.1097</i>	ALS family protein with roles in adhesion, biofilm formation, germ tube induction	Adhesion	Wh		Wh	5.5	1.4	1.0	4.6	1.0	0.2*	0.2*	1.0
ADH1 <i>orf19.3997</i>	Alcohol dehydrogenase	Adhesion			Wh	20.9	4.7	1.0	21.7	2.6	0.0*	0.1*	0.4
<i>orf19.4216</i>	<i>Putative heat shock protein</i>	Adhesion			Wh	37.1	28.2	3.1	38.9	1.0	0.1*	0.0*	3.1
CSH1 <i>orf19.4477</i>	Aldo-keto reductase family member with role in fibronectin adhesion	Adhesion			Wh	2.2	2.0	1.0	2.4	1.0	0.5*	0.4*	1.0
ALS4 <i>orf19.4555</i>	ALS family protein with roles in adhesion and germ tube induction	Adhesion	Wh	Wh	Wh	8.4	2.0	1.0	7.6	1.2	0.1*	0.2*	0.8
EFG1 <i>orf19.610</i>	Transcriptional repressor required for white-phase cell type, hyphal growth, metabolism with roles in adhesion and virulence	Adhesion	Wh	Wh	Wh	5.3	2.0	1.1	4.8	1.0	0.2*	0.2*	1.1

DOWNREGULATED IN GUT ONLY (VS. WH CONTROLS)													
FRE10 orf19.1415	Major cell-surface ferric reductase	Iron uptake			Wh	11.0	5.1	1.0	9.6	2.9	0.1*	0.3*	0.3*
CFL4 orf19.1932	Putative ferric reductase	Iron uptake			Wh	7.0	2.0	1.0	9.1	2.0	0.1*	0.2	0.5
SIT1 orf19.2179	Transporter of ferrichrome siderophores	Iron uptake		Wh		2.1	1.9	1.0	1.9	1.6	0.5*	0.8	0.6
FET34 orf19.4215	Putative multicopper ferroxidase	Iron uptake	Wh		Wh	29.0	2.1	1.0	23.7	1.3	0.0*	0.1	0.8
FTH1 orf19.4802	Protein similar to <i>S. cerevisiae</i> Fth1p, a high affinity iron transporter	Iron uptake			Wh	2.1	1.2	1.0	2.0	1.4	0.5*	0.7	0.7
FTR1 orf19.7219	High-affinity iron permease	Iron uptake			Wh	7.1	1.9	1.0	6.2	1.8	0.1*	0.3*	0.5*
FTR2 orf19.7231	High-affinity iron permease	Iron uptake		Op		2.8	1.3	1.0	2.4	1.5	0.4*	0.6	0.7*
PDC11 orf19.2877	Putative pyruvate decarboxylase	Glucose Utilization				10.1	3.9	1.0	7.0	1.9	0.1*	0.3	0.5
PGK1 orf19.3651	Phosphoglycerate kinase	Glucose Utilization		Wh		2.1	1.6	1.0	1.8	1.6	0.5*	0.9	0.6*
PFK1 orf19.3967	Phosphofructokinase alpha subunit	Glucose Utilization		Wh		2.1	1.5	1.0	2.0	1.5	0.5*	0.8	0.7*
ADH1 orf19.3997	Alcohol dehydrogenase	Glucose Utilization			Wh	20.9	4.7	1.0	21.7	2.6	0.0*	0.1*	0.4
FBA1 orf19.4618	Putative fructose-bisphosphate aldolase	Glucose Utilization			Wh	2.2	1.4	1.0	1.7	1.4	0.5*	0.9	0.7
ADH2 orf19.5113	Alcohol dehydrogenase	Glucose Utilization	Op	Wh	Op	11.8	3.1	1.0	9.6	2.6	0.1*	0.3*	0.4
HXK2 orf19.542	Hexokinase II	Glucose Utilization		Wh		2.1	1.5	1.0	1.9	1.6	0.5*	0.8	0.6*

HIGHER IN OPAQUE THAN GUT (DIRECT COMPARISON, IN ADDITION TO SAP GENES)													
STE4 <i>orf19.799</i>	Beta subunit of heterotrimeric G protein required for mating	Mating			Op	1.2	1.5	1.0	3.7	15.9	0.9	4.3*	0.1*
CAG1 <i>orf19.4015</i>	Alpha subunit of heterotrimeric G protein required for mating	Mating				1.0	1.3	1.2	14.2	20.7	1.2	1.5	0.1*
STE18 <i>orf19.6551.1</i>	Gamma subunit of heterotrimeric G protein involved in mating	Mating				1.0	1.1	1.7	12.1	15.2	1.7	1.3	0.1*
STE2 <i>orf19.696</i>	Receptor for alpha factor mating pheromone	Mating				1.1	1.2	1.0	3.7	4.0	0.9	1.1	0.2*
HIGHER IN GUT THAN OPAQUE													
GIT2 <i>orf19.1978</i>	glycerophosphoinositol permease	Phosphate transport			Wh	3.6	2.9	2.2	4.3	1.0	0.6*	0.2*	2.2*
GIT3 <i>orf19.1979</i>	<i>Putative glycerophosphoinositol permease</i>	Phosphate transport	Wh	Wh	Wh	28.1	8.0	2.8	26.7	1.0	0.1*	0.0*	2.8*
PHO84 <i>orf19.655</i>	High-affinity phosphate transporter	Phosphate transport	Op		Op	1.3	4.7	14.4	1.0	4.2	10.7*	4.2*	3.5*
<i>orf19.6888</i>	Putative transcription factor with Gal4p-like DNA-binding domain	Transcription Factor			Wh	2.0	1.9	2.9	1.9	1.0	1.5	0.5*	2.9*
<i>orf19.4914</i>	Putative transcription factor	Transcription Factor			Wh	2.1	1.8	2.8	2.2	1.0	1.3	0.5*	2.8*
LEU3 <i>orf19.4225</i>	Putative transcription factor with zinc-finger domain	Transcription Factor				3.8	3.4	3.2	1.1	1.0	0.8	0.9	3.2*
<i>orf19.2726</i>	Putative plasma membrane protein	Cell Wall or Secreted	Wh	Wh	Wh	10.2	6.6	4.3	9.2	1.0	0.4*	0.1*	4.3*
<i>orf19.5070</i>	Putative cell-wall mannoprotein	Cell Wall or Secreted	Wh			1.8	2.0	3.0	1.7	1.0	1.7*	0.6	3.0*
<i>orf19.1258</i>	Putative adhesin-like protein	Cell Wall or Secreted	Wh	Wh	Wh	6.3	6.9	2.9	6.5	1.0	0.5*	0.2*	2.9*
RBT4 <i>orf19.6202</i>	Putative secreted protein that is required for virulence	Cell Wall or Secreted			Wh	7.1	3.8	2.7	6.7	1.0	0.4*	0.1*	2.7*

RBE1 <i>orf19.7218</i>	Cell wall protein	Cell Wall or Secreted				1.3	1.6	2.4	1.1	1.0	1.9*	0.9	2.4*
<i>orf19.5267</i>	Putative cell wall adhesin-like protein	Cell Wall or Secreted			Op	1.2	1.3	5.6	1.0	2.2	4.9*	2.2*	2.6*
RHD1 <i>orf19.54</i>	Putative beta-mannosyltransferase	Cell Wall Structure	Wh	Wh	Wh	7.1	4.0	4.8	6.5	1.0	0.7	0.2*	4.8*
CIS2 <i>orf19.6053</i>	Putative role in regulation of biogenesis of the cell wall	Cell Wall Structure			Wh	2.3	1.9	2.4	2.4	1.0	1.1	0.4*	2.4*
SKN2 <i>orf19.348</i>	Protein with a potential role in beta-1,6 glucan biosynthesis	Cell Wall Structure			Wh	2.5	1.6	2.4	2.9	1.0	1.0	0.3*	2.4*

Descriptions of published white (w) and opaque (o) gene sets refer to: ²⁸ Lan et al.(2002) PNAS 23: 14902-12; ⁴³ Tsong et al. (2003) Cell 115: 389-99; ⁴⁴ Tuch et al. (2010) PLoS Genetics 6: e1001070

Supplementary Table 5. Strains Used in this Study

Strain	Nickname	MTL genotype	Phase	Full Genotype	Parent	Reference
SN78	Wild type	MTLa/α	white	<i>leu2Δ/leu2Δ, ura3Δ/ura3Δ, his1Δ/his1Δ, iro1Δ/iro1Δ, MTLa/MTLα</i>	RM1000 #2	14
SN87	Wild type	MTLa/α	white	<i>leu2Δ/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, iro1Δ/IRO1, MTLa/MTLα</i>	RM1000 #2	14
SN152	Wild type	MTLa/α	white	<i>leu2Δ/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLa/MTLα</i>	RM1000 #2	14
SN235	Wild type	MTLa/α	white	<i>leu2Δ::C.d.HIS1/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLa/MTLα</i>	SN152	This study
SN250	Wild type	MTLa/α	white	<i>leu2Δ::C.m.LEU2/leu2Δ::C.d.HIS1, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLa/MTLα</i>	SN152	42
SN425	Wild type	MTLa/α	white	<i>leu2Δ::C.d.HIS1/leu2Δ::C.m.LEU2, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ::C.d.ARG4, iro1Δ/IRO1, MTLa/MTLα</i>	SN152	42
QMY23	Wild type	MTLa/α	white	<i>leu2Δ::C.m.LEU2/leu2Δ::C.d.HIS1, ura3Δ/URA3, his1Δ/his1Δ, iro1Δ/IRO1, MTLa/MTLα</i>	SN87	39
SN1011	efg1Δ/Δ	MTLa/α	white	<i>efg1Δ::C.m.LEU2/efg1Δ::C.d.HIS1, leu2Δ/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLa/MTLα</i>	SN152	14
SN999	wor1Δ/ WOR1	MTLa/α	white	<i>WOR1/wor1Δ::C.d.HIS1, leu2Δ/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLa/MTLα</i>	SN152	This study
SN881	wor1Δ/Δ	MTLa/α	white	<i>wor1Δ::C.m.LEU2/wor1Δ::C.d.HIS1, leu2Δ/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLa/MTLα</i>	SN152	This study
RZY244	wor1Δ/Δ	MTLa/α	white	<i>wor1Δ::C.m.LEU2/wor1Δ::C.d.HIS1, leu2Δ/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, iro1Δ/IRO1, MTLa/MTLα</i>	SN87	Zordan & Johnson, un-published
SN1064	wor1Δ/Δ	MTLa/α	white	<i>wor1Δ::C.m.LEU2/wor1Δ::C.d.HIS1, leu2Δ/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/ARG4, iro1Δ/IRO1, MTLa/MTLα</i>	SN152	This study
SN1014	WOR1 _{prom} -FLP intermediate	MTLa/α	white	<i>wor1Δ::FLP-SAT1/WOR1, leu2Δ/leu2Δ, ura3Δ/ura3Δ, his1Δ/his1Δ, iro1Δ/iro1Δ, MTLa/MTLα</i>	SN78	This study
SN1020	WOR1 _{prom} -FLP	MTLa/α	white	<i>wor1Δ::FLP-SAT1/WOR1, leu2Δ::FRT-URA3-FRT/leu2Δ, ura3Δ/ura3Δ, his1Δ/his1Δ, iro1Δ/iro1Δ, MTLa/MTLα</i>	SN1014	This study
SN967*	MTLa opaque	MTLa/a	opaque	<i>leu2Δ::C.m.LEU2/leu2Δ::C.d.HIS1, ura3Δ/URA3, his1Δ/his1Δ, iro1Δ/IRO1, MTLa/MTLα</i>	QMY23	35
SN966	MTLa	MTLa/a	white	<i>leu2Δ::C.m.LEU2/leu2Δ::C.d.HIS1, ura3Δ/URA3, his1Δ/his1Δ, iro1Δ/IRO1, MTLa/MTLα</i>	SN967	This study
SN1008	MTLα opaque	MTLΔ/α	opaque	<i>leu2Δ/leu2Δ::SAT1, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1,</i>	SN152	This study

				<i>mtlαΔ::C.d.HIS1/MTLα</i>		
SN1038	<i>MTLα</i> opaque	<i>MTLα/Δ</i>	opaque	<i>leu2Δ/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLα/mltαΔ::C.d.ARG4</i>	SN152	This study
SN927	<i>WOR1^{OE}/WOR1</i>	<i>MTLα/α</i>	white	<i>SAT1-TDH3_{prom}-WOR1/WOR1, leu2Δ::C.m.LEU2/leu2Δ::C.d.HIS1, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLα/MTLα</i>	SN250	This study
SN928	<i>WOR1^{OE}</i>	<i>MTLα/α</i>	white	<i>SAT1-TDH3_{prom}-WOR1/wor1Δ::C.d.ARG4, leu2Δ::C.m.LEU2/leu2Δ::C.d.HIS1, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLα/MTLα</i>	SN927	This study
SN1001	<i>WOR1^{OE}</i>	<i>MTLα/α</i>	white	<i>SAT1-TDH3_{prom}-WOR1/wor1Δ::C.d.HIS1, leu2Δ/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, iro1Δ/IRO1, MTLα/MTLα</i>	SN999	This study
SN1044	<i>WOR1^{OE}</i> (post-mouse)	<i>MTLα/α</i>	white	<i>SAT1-TDH3_{prom}-WOR1/wor1Δ::C.d.ARG4, leu2Δ::C.m.LEU2/leu2Δ::C.d.HIS1, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLα/MTLα</i>	SN928	This study
SN1045	<i>WOR1^{OE}</i> GUT (post-mouse)	<i>MTLα/α</i>	GUT	<i>SAT1-TDH3_{prom}-WOR1/wor1Δ::C.d.ARG4, leu2Δ::C.m.LEU2/leu2Δ::C.d.HIS1, ura3Δ/URA3, his1Δ/his1Δ, arg4Δ/arg4Δ, iro1Δ/IRO1, MTLα/MTLα</i>	SN928	This study
SN1046	<i>WOR1^{OE}</i> GUT (post-mouse)	<i>MTLα/α</i>	GUT	<i>SAT1-TDH3_{prom}-WOR1/wor1Δ::C.d.HIS1, leu2Δ/leu2Δ, ura3Δ/URA3, his1Δ/his1Δ, iro1Δ/IRO1, MTLα/MTLα</i>	SN1001	This study

C.d. ARG4 denotes *Candida dubliniensis* *ARG4*, *C.d. HIS1* denotes *Candida dubliniensis* *HIS1*, and *C.m.*

LEU2 denotes *Candida maltosa* *LEU2*.

* SN967 was constructed by Mitrovich et al.³⁵ but was not assigned a unique name to distinguish it from its *MTLα/α* parent (QMY23).

Supplementary Table 6. Plasmids Used in this Study

Plasmid	Insert	Use	Selectable Markers	Vector
pSN209	<i>Pmel</i> -sequence ending 120 bp upstream of <i>WOR1</i> ORF- <i>SAT1</i> gene- <i>TDH3</i> promoter-1 st 300 bp of <i>WOR1</i> ORF- <i>Pmel</i>	Replacement of <i>WOR1</i> promoter with (<i>SAT1</i> gene and) <i>TDH3</i> promoter	AmpR, <i>ScURA3</i> , NatR	pRS316
pSN288	<i>Pmel</i> -final 500 bp of <i>WOR1</i> promoter- <i>FLP</i> ORF- <i>SAT1</i> gene- <i>WOR1</i> downstream sequence- <i>Pmel</i>	Replacement of <i>WOR1</i> ORF with <i>FLP</i> ORF	AmpR, <i>ScURA3</i> , NatR	pRS316
pSN290	<i>Pmel</i> - <i>LEU2</i> upstream sequence-FRT- <i>C. albicans URA3</i> -FRT- <i>LEU2</i> downstream sequence- <i>Pmel</i>	Introduction of FRT- <i>URA3</i> -FRT at the <i>LEU2</i> locus	AmpR, <i>ScURA</i>	pRS316

Pmel denotes the recognition sequence for this restriction enzyme; *ScURA3* denotes the *URA3* gene from *Saccharomyces cerevisiae*, AmpR denotes resistance to ampicillin, NatR denotes resistance to nourseothricin.

Supplementary Table 7. Primers Used in this Study

Primer Name	Purpose	Sequence (5' to 3')
SNO1122	wor1-ko gene disruption primer 5' flank, forward (SN881 and SN1064)	TCCGTATTGGTATTGGTAG
SNO1123	wor1-ko gene disruption primer 5' flank, reverse (SN881 and SN1064)	CACGGCGCGCCTAGCAGCGGTAAGACAGATATTGAAACTTCAC
SNO1124	wor1-ko gene disruption primer 3' flank, forward (SN881 and SN1064)	GTCAGCGGCCGCATCCCTGCATCTTATATATGTGGTCTGTG
SNO1125	wor1-ko gene disruption primer 3' flank, reverse (SN881 and SN1064)	TTCAGCTCTCTAAGTAGTATTG
SNO1271	Common primer for amplifying His, Leu and Arg cassettes, forward	CCGCTGCTAGGCGGCCGTGACCAGTGTGATGGATATCTGC
SNO1272	Common primer for amplifying His, Leu and Arg cassettes, reverse	GCAGGGATGCGGCCGTGACAGCTCGGATCCAATAGTAACG
SNO1126	Wor1 ORF check primer, forward	GACGACGAGTACGACCAACAA
SNO1127	Wor1 ORF check primer, reverse	CAGCTTCCCTCCATGTGT
SNO1128	Wor1 integration verification check left, forward	TTTCTTCCAAAACCTGCC
SNO1129	Wor1 integration verification check right, reverse	TCCCTTCATGAATAGTTCCC
SNO1130	<i>C. dubliniensis HIS1</i> integration verification check left, reverse	ATTAGATACTGGTGGTTC
SNO1131	<i>C. dubliniensis HIS1</i> integration verification check right, forward	AACACAAC TGACAATCTGG
SNO1132	<i>C. maltosa LEU2</i> integration verification check left, reverse	AGAATTCCCAACTTGTCTG
SNO1133	<i>C. maltosa LEU2</i> integration verification check right, forward	AAACTTGAAACCGGCTGCG
SNO143	<i>C. dubliniensis ARG4</i> integration on <i>C. albicans</i> ARG4 locus, 5' flank, forward	GGACAGAAAGTTATTGTACAG
SNO144	<i>C. dubliniensis ARG4</i> integration on <i>C. albicans</i> ARG4 locus, 5' flank, reverse	TCACGGCGCGCCTAGCAGCGGATTATTCTTGTAGCTGTTATG
SNO1385	<i>C. dubliniensis ARG4</i> integration on <i>C. albicans</i> ARG4 locus, ARG4 insert, forward	CCGCTGCTAGGCGGCCGTGAAACGAATCAGACTCTGATACCCAGTGTGATGGATATCTGC
SNO241	<i>C. dubliniensis ARG4</i> integration on <i>C. albicans</i> ARG4 locus, ARG4 insert, reverse	GCAGGGATGCGGCCGTGACAGCTCGGATCCACATAGTAACG
SNO145	<i>C. dubliniensis ARG4</i> integration on <i>C. albicans</i> ARG4 locus, 3' flank, forward	GTCAGCGGCCGCATCCCTGCCTCATATAATAATCACAGTATTG
SNO146	<i>C. dubliniensis ARG4</i> integration on <i>C. albicans</i> ARG4 locus, 3' flank, reverse	TCAGACGATCTTACAATGG
SNO187	<i>C. dubliniensis ARG4</i> integration verification check, left, forward	CAAGAGTAGTCTCAAATAAAC
SNO263	<i>C. dubliniensis ARG4</i> integration verification check, left, reverse, qPCR for wor1ko (SN1064), reverse	TTCAACCTTCAAACGATGC
SNO264	<i>C. dubliniensis ARG4</i> integration verification check right, forward	TCGATACATTGCGGTACAG
SNO188	<i>C. dubliniensis ARG4</i> integration verification check, right, reverse	CGTTTGGAAAGCTGTATATCG
SNO183	qPCR for SN425 WT, SN250 WT and QMY23 WT, forward	GATGCCTTAGCTCATTCTC
SNO322	Common qPCR primer for SN425 WT, SN250 WT, SN235 WT, efg1ko and wor1ko (SN881), reverse	TGAGCACCATAAGGACGTTCTC

SNO1003	Common qPCR primer for QMY23 WT and <i>wor1ko</i> (RZY244), reverse	CCGGTTTACTTGGATCTTCG
SNO1007	qPCR for <i>wor1ko</i> (RZY244), forward	GTACACTGACATCTCAAACATCAA
ST49	qPCR for <i>wor1ko</i> (SN1064), forward	AACGAATCAGACTCTGATAC
STO8	qPCR for <i>efg1ko</i> , forward	TTTGTGGAGCCTTCATGA
STO2	qPCR for <i>wor1ko</i> (SN881) and <i>wor1het</i> (SN999), forward	AAGATTGGTTCTGTCGACA
SNO1697	qPCR for <i>wor1het</i> (SN999), reverse	CGTAGCCATGAGCACCAAA
SNO1134	<i>WOR1</i> -OE and <i>WOR1p-FLP</i> strain construction; amplifies <i>WOR1</i> upstream region, forward	GGCGAATTGGAGCTCCACCGCGGTGGCGGCCGCTCTAGAACTAGTGGATCGTTAACCTTCCAA AACCCTGCCTT
SNO1135	<i>WOR1</i> -OE strain construction; amplifies <i>WOR1</i> upstream region, reverse	GGGACGAGGCAGCTGATTCTTGATGTTGAGATGTCAGTG
SNO1136	<i>WOR1</i> -OE strain construction; amplifies <i>SAT1</i> cassette, forward	CTGACATCTCAAACATCAAAGAACATCAAGCTTGCC TCGTCCCC
SNO1137	<i>WOR1</i> -OE strain construction; amplifies <i>SAT1</i> cassette, reverse	GACTATACTGAATTAGACATATTGAATTCAATT GTGATG
SNO1138	<i>WOR1</i> -OE strain construction; amplifies <i>WOR1</i> ORF fragment, forward	CATCACAAATTGAATTCAAATATGTCTAATTCAAGT ATAGTCC
SNO1139	<i>WOR1</i> -OE strain construction; amplifies <i>WOR1</i> ORF fragment, reverse	GTCGACGGTATCGATAAGCTGATATCGAATTCC TGCAGCCCGGGGATCGTTAACCTGTGGTCG TACTCGTCGTCGGGACC
SNO1187	<i>WOR1</i> -OE 5' integration verification check, forward	CCTATTGTTATTGCAGAACAG
SNO1188	<i>WOR1</i> -OE 5' integration verification check, reverse	GTATTCTGGGCCTCCATGTC
SNO1185	<i>WOR1</i> -OE 3' integration verification check, forward	AACCCTTGAAATTCCCTTCA
SNO1186	<i>WOR1</i> -OE 3' integration verification check, reverse	GTGGTTACCATACCCACCAG
SNO1342	qPCR for SN425 WT, reverse	TCAAACGAAGGTACACTGACT
SNO1343	qPCR for SN425 WT, forward	AACAGCTATCAAGAATAATCCGC
STO50	alternate qPCR for SN425 WT, forward	TATGCAGAGAGATATACATC
SNO1355	qPCR for <i>WOR1</i> -OE, forward	CCAATTGAAAGACCATTACGC
SNO1361	qPCR for <i>WOR1</i> -OE, reverse	GCACGTCAAGACTGTCAAGGAGG
SNO1521	qPCR for SN967 WT, forward	TCTAGTGGTGAATTGGGTT
SNO1522	qPCR for SN967 WT, reverse	TAAGTAGATGGAATAGATTGATC
SNO1008	PCR check for the presence of <i>OBPa</i> , forward	GCATATTGCACCAAGGCAG
SNO1009	PCR check for the presence of <i>OBPa</i> , reverse	GATTCCATGACCTCGTTCC
SNO1010	PCR check for the presence of <i>OBPalpha</i> , forward	GAAGATGACTCAGGTCATGC
SNO1011	PCR check for the presence of <i>OBPalpha</i> , reverse	CTTCAATTGCATCGTAAGTACC
SNO1531	<i>WOR1p-FLP</i> strain construction; amplifies <i>WOR1</i> upstream region, reverse	CATAATATCAAATTGTGGCATTGCTTAATATTG AATTGAATTATAC
SNO1532	<i>WOR1p-FLP</i> strain construction; amplifies <i>FLP-SAT1</i> cassette, forward	GTATAATTCAATTCAATATTAAGCAATGCCACAATTGATATATTATG
SNO1533	<i>WOR1p-FLP</i> strain construction; amplifies <i>FLP-SAT1</i> cassette, reverse	CCTGAATGAGCCCCAAAATAAACAGGGACCACC TTGATTGTAAA
SNO1534	<i>WOR1p-FLP</i> strain construction; amplifies ~450bp of <i>WOR1</i> 3'UTR, forward	TTTACAATCAAAGGTGGCCTGTTATTATTTGGG GCTCATTCAAGG
SNO1352	<i>WOR1p-FLP</i> strain construction; amplifies ~450bp of <i>WOR1</i> 3'UTR, reverse	GTCGACGGTATCGATAAGCTGATATCGAATTCC TGCAGCCCGGGGATCGTTAACCGTCAAGGGG TCATCATATCATT
SNO1535	<i>WOR1p-FLP</i> strain verification check, left,	AGAAAGAAAGAGAGAGAGGAAACG

	forward	
SNO1536	<i>WOR1p-FLP</i> strain verification check, left, reverse	CTGTTCCGTTATGTGAATCATCC
SNO1537	<i>WOR1p-FLP</i> strain verification check, right forward	CGCCTAACATATGTGAAGTGTGA
SNO1354	<i>WOR1p-FLP</i> strain verification check, right, reverse	CGTTCAGATATTACATACATCCACCT
SNO464	<i>Leu2-FRT-URA-FRT</i> strain construction; amplifies <i>Leu2</i> 5' flank, forward	GCGAATTGGAGCTCCACCGCGGTGGCGGCCG CTCTAGAACTAGTGGATCGTTAAACTGGTAGA TTTACAACCTGAAGCCG
SNO1538	<i>Leu2-FRT-URA-FRT</i> strain construction; amplifies <i>Leu2</i> 5' flank, reverse	CAGTAGCTCGAGTTAAATCCGAAGTTCCTATTCT CTAGAAAGTATAGGAACCTCCTCGAGGGAAAAGG GGAGTATTCTGGAGTGAA
SNO1539	<i>Leu2-FRT-URA-FRT</i> strain construction; amplifies <i>C.a. URA3</i> cassette, forward	AGAAATACTCCCCTTCTCGAGGAAGTTCCTA TACTTCTAGAGAATAGGAACCTCGGATTTAACTC GAGCTACTGATATCAATGC
SNO1540	<i>Leu2-FRT-URA-FRT</i> strain construction; amplifies <i>C.a. URA3</i> cassette, reverse	GCTAACTACTGTATATACTGGGATCTGAAGTTC TATTCTCTAGAAAGTATAGGAACCTCCTGCTTAA TCGATATAACTGGTTGA
SNO1541	<i>Leu2-FRT-URA-FRT</i> strain construction; amplifies <i>Leu2</i> 3' flank, forward	CAAGTTATATCGATTAAGCAGGAAGTTCCTATA CTTTCTAGAGAATAGGAACCTCAGATCCCAGTAT ATACAGTAGTTAGCATTAA
SNO467	<i>Leu2-FRT-URA-FRT</i> strain construction; amplifies <i>Leu2</i> 3' flank, reverse	GTCGACGGTATCGATAAGCTGATATCGAATTCC TGCAGCCGGGGATCGTTAAACTCGAAAACG ATGTTGCACCACCG
SNO1443	<i>Leu2-FRT-URA-FRT</i> strain verification check, left. forward	GCTTGAGTTCTGGGTCA
SNO1547	<i>Leu2-FRT-URA-FRT</i> strain verification check, left. reverse	GTTGACCGTATCAGTAGCATCATC
SNO1447	<i>Leu2-FRT-URA-FRT</i> strain verification check, right. reverse	CGCTACCCAACAAAGAGACC
SNO1542	<i>Leu2-FRT-URA-FRT</i> strain verification check, right. forward	GCAATCGAAGTAGCTGGATA
SNO509	URA excision verification primer, and qPCR for <i>SN235 WT</i> , forward	GTTGTGATTTGCTATTCCGGCGCT
SNO840	URA excision verification primer, reverse	TCTCTCCGAATGAAGAGCC
SNO1650	<i>Leu2</i> knockout verification primer for strains <i>SN78</i> and <i>SN1020</i> , forward	GAAATGCTGGTTGGAATGCT
SNO1652	<i>Leu2</i> knockout verification primer for strains <i>SN78</i> and <i>SN1020</i> , reverse	GCGGTCTAGAAGGACCACCT
SNO819	qRTPCR for <i>ACT1</i> ORF, forward	GTGGTACTACCATGTTCCCAGG
SNO820	qRTPCR for <i>ACT1</i> ORF, reverse	GATAGAACCAATCCAGACAGAG
SNO1154	qRTPCR for <i>WOR1</i> ORF, forward	TGCTGGTGGATCTAGTAGTGTAGC
SNO1155	qRTPCR for <i>WOR1</i> ORF, reverse	AGTACCGGTGTAATACGACCCAGA
SNO1603	qRTPCR for <i>TDH3</i> ORF, forward	GCTCCAGACTATGCTGCTTACAT
SNO1604	qRTPCR for <i>TDH3</i> ORF, reverse	GGAATGTTAGCTGGGTCTTTTC
SNO1332	<i>MTLa-ko</i> gene disruption primer 3' flank, forward	GGAATTATTGTCTAACCTGCCGTGG
SNO1333	<i>MTLa-ko</i> gene disruption primer 3' flank, reverse	CGTTACTAGTGGATCCGAGCTGCCGCATAT ACACTGGAAATAAGTGGTGGTAGT
SNO1336	<i>MTLa-ko</i> gene disruption primer 5' flank, forward	TCAATCAGGTGCGGTGTGGCGCGCCTATATGT ACATACAAAATGGTTATTGTAGCAGG
SNO1335	<i>MTLa-ko</i> gene disruption primer 5' flank, reverse	CCCTTGGGTAGTAAAGGTAAAGCCAATGCCG
SNO1315	<i>MTLa-ko</i> gene disruption, primer to amplify <i>C. dubliniensis HIS1</i> cassette, forward	TATATGCCGCCGCAGCTGGATCCACTAGTAAC G

SNO1318	<i>MTLa</i> -ko gene disruption, primer to amplify <i>C. dubliniensis</i> <i>HIS1</i> cassette, reverse	ATATAGGCGGCCACACCGAACCTGATTGATAC CAGTGTGATGGATATCTGC
SNO1337	<i>MTLa</i> -ko 3' integration verification check, reverse	GTTGGTGAACCTAAAGTACTTTATTGG
SNO1339	<i>MTLa</i> -ko 3' integration verification check, forward	AGTGGATCCGAGCTCGGGCCGCATATA
SNO1340	<i>MTLa</i> -ko 5' integration verification check, forward	TTGAGTGTTGCTGGTCTTGC
SNO1341	<i>MTLa</i> -ko 5' integration verification check, reverse	TGCAGATATCCATCACACTGG